The results presented in Table 1 show that the incorporation of 1 into 2 and 3 in V. offinalis was extremely efficient in young shoots. During the flowering period, however, the incorporation decreased significantly. Previously, we had shown [3] that the incorporation of 1 into 3 in V.

officinalis plants, past the flowering stage, was too small to be measured by 2H NMR (< 0.5 %).

These results suggest that immature plants should be used in biosynthetic experiments with advanced precursors, unless other information is at hand.

EXPERIMENTAL

The experimental techniques, as well as the synthesis and physical properties of the precursor, have been described previously [1].

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REFERENCES

- Damtoft, S., Jars, M. U., Kirk, O., Jensen, S. R. and Nielsen, B. J., (1983) Phytochemistry 22, 695.
- Horodysky, A. G., Waller, G. R. and Eisenbraun, E. J. (1969) J. Biol. Chem. 244, 3110.
- Damtoft, S. (1980) Thesis, The Technical University of Denmark, Lyngby.

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GUAIANOLIDES FROM CENTAUREA KOTSCHYI

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Key Word Index—Centaurea kotschyi; Compositae; sesquiterpene lactones; guaianolides.

Abstract—The aerial parts of Centaurea kotschyi (var. kotschyi) afforded, besides three known sesquiterpene lactones, a new derivative of linichlorin B. The structure was determined by spectral analysis.

INTRODUCTION

In continuation of our phytochemical investigation of the genus *Centaurea* [1], we report here the isolation and structure elucidation of a new guaianolide.

Centaurea kotschyi Boiss., endemic to Turkey, had not been investigated chemically. The aerial parts of the plant gave, in addition to the three known guaianolides desacylcynaropicrin (1) [2], cynaropicrin (2) [3] and linichlorin B (3) [4], a new lactone (4) which is closely related to linichlorin B.

RESULTS AND DISCUSSION

Compound 4, $C_{19}H_{24}O_7$, [M]⁺ 364, exhibited the typical IR absorption bands of an α,β -unsaturated- γ -lactone group (1750 cm⁻¹), hydroxyl group(s) (3400 cm⁻¹)

and an acyl group (1730 and 1270 cm⁻¹). The ¹H NMR spectrum (Table 1) showed that compound 4 was very similar to linichlorin B, and that they both had the same ring structure, therefore the only difference was in the side chain. A pair of doublets at δ 3.87 and 3.64 (J=11 Hz) indicated the presence of a -CH₂-O- group [5, 6] and a methyl singlet δ 1.40, in addition to the signals that were assigned for the ring protons, suggested that the acyl group was an α , β -dihydroxyisobutyryl moiety. In the mass spectrum peaks at m/z 244 [M-C₄H₈O₄]⁺ and 119 corroborated the presence of this group.

EXPERIMENTAL

Extraction and isolation. The dried, aerial parts of C. kotschyi (var. kotschyi) (1.5 kg), collected near Ankara in July 1980 (voucher No. 45503 deposited at the herbarium of the Faculty of

$$1 R = H$$

$$2 R = OH$$

$$OH$$

$$OH$$

$$OH$$

Pharmacy, Istanbul), were extracted with EtOH, evapd to small vol. *in vacuo*, and worked up in the usual manner by using an aq. soln of 4% lead acetate [1]. The residue (13.6 g) was chromatographed on a silica gel column and eluted with CHCl₃ containing increasing amounts of MeOH. The fractions yielded 1, 2, 3 and 4 (90, 25, 50 and 17 mg), respectively.

Compound 4. This product was purified by prep. TLC; IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3400 (OH), 1750 (γ -lactone), 1730 and 1270 (CO₂R), 1635 (double bond); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 215; MS m/z (rel. int.): C₁₉H₂₄O₇, 364 [M]⁺ (7.0), 261 [M-C₄H₇O₃]⁺ (11.4), 244 [M-C₄H₈O₄]⁺ (36.9), 119 (35), 91 (74.8) 75 (100).

REFERENCES

1. Oksuz, S., Ulubelen, A., Aynechi, Y. and Wagner, H. (1982) *Phytochemistry* 21, 2747.

Table 1. ¹H NMR spectral data of 4 (400 MHz, CHCl₃, TMS as internal standard)

Н	δ
1α	2.99 ddd
2α	1.71 ddd
2β	2.21 ddd
3α	4.57 t(br)
5α	$2.81 \ t(br)$
6β	4.28 dd (10.5; 9)*
7α	3.18 dddd
8β	5.15 ddd (5.5; 4; 9.5)
9α	2.68 dd (5.5; 14.5)
9β	2.47 dd (3.5; 15)
13	6.24 d (3.4)
13'	5.60 d (3)
14	$5.16 \ s \ (br)$
14'	$5.09 \ s(br)$
15	$5.50 \ s(br)$
15'	$5.37 \ s(br)$
-CCH ₂ -OH	3.87 d (11)
-	3.64 d (11)
-C(OH)CH ₃	1.40 s

- *J (Hz).
- Gonzales, A. G., Bermejo, J., Massanet, G. M. and Perez, J. (1973) An. Quim. 69, 1333.
- 3. Rustaiyan, A., Niknejad, A., Zdero, C. and Bohlmann, F. (1981) *Phytochemistry* **20**, 2427.
- Gonzales, A. G., Bermejo, J., Amaro, J. M., Massanet, G. M., Galindo, A. and Cabrera, I. (1978) Can. J. Chem. 56, 491.
- Gonzales, A. G., Bermejo, J., Breton, L., Massanet, G. M. and Triana, J. (1974) Phytochemistry 13, 1193.
- Suchy, M., Samek, Z., Herout, V. and Sorm, F. (1968) Collect. Czech. Chem. Commun. 33, 2238.